

Chemical Communications

Number 17
1991

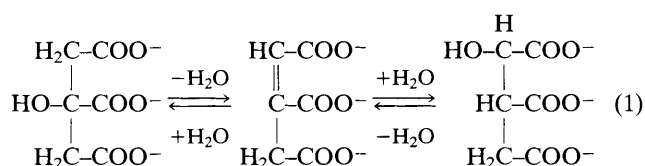
Kinetic Studies on the Two-stage Activation of Aconitase

K. Y. Faridooon, Hua-Yun Zhuang and A. Geoffrey Sykes*

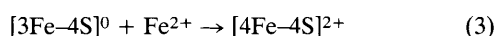
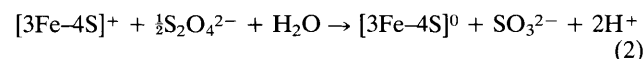
Department of Chemistry, The University, Newcastle upon Tyne NE1 7RU, UK

Kinetic studies on the stepwise activation of aconitase $[3\text{Fe-4S}]^+ \rightarrow [3\text{Fe-4S}]^0$ (reduction), followed by $[3\text{Fe-4S}]^0 \rightarrow [4\text{Fe-4S}]^{2+}$ (Fe^{2+} incorporation) are described.

The enzyme aconitase catalyses the dehydration–rehydration process resulting in the interconversion of citrate and *iso*-citrate *via* the allylic intermediate *cis*-aconitate [eqn. (1)], a reaction which is a part of the Kreb's cycle.¹

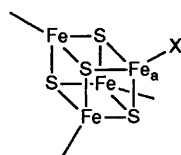


In this work aconitase was isolated from fresh beef hearts using a modified form of existing procedures.²⁻⁵ The purity of the protein was estimated to be >95% by comparing its activity with literature values.⁵ The crystal structure of pig heart aconitase has been refined to 2.5 Å resolution.⁶ Active aconitase has a $[4\text{Fe-4S}]^{2+}$ cuboidal cluster close to the centre of the molecule ($M_r \sim 80,000$; 755 amino acids), which is accessed by a cleft.⁷ One of the Fe's in the cluster designated Fe_a , coordinated by X (either H_2O or OH^-) is labile. In the course of the aerobic isolation Fe_a is lost and the inactive $[3\text{Fe-4S}]^+$ containing enzyme is obtained.⁵ *In vitro* activation can be achieved by reducing $[3\text{Fe-4S}]^+$ to $[3\text{Fe-4S}]^0$ with dithionite, followed by Fe^{2+} incorporation to give $[4\text{Fe-4S}]^{2+}$ [eqns. (2) and (3)].



The different states of aconitase are shown in Scheme 1. All the forms indicated except $[4\text{Fe-4S}]^{3+}$ have been identified. Approximate reduction potentials for the $[3\text{Fe-4S}]^+/[3\text{Fe-4S}]^0$ (-0.16 V) and $[4\text{Fe-4S}]^{2+}/[4\text{Fe-4S}]^+$ (-0.50 V) couples have been indicated.

Here we report for the first time kinetic studies relating to the activation process, including the trimer \rightarrow cube conversion. Relevant spectra are indicated in Fig. 1.⁸ Samples of inactive $[3\text{Fe-4S}]^+$ enzyme were made air-free by (Amicon) diafiltration using air-free solutions. These and dithionite



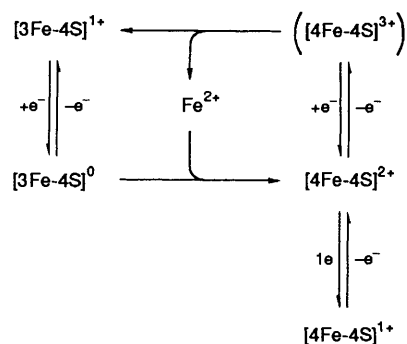
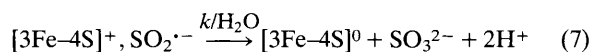
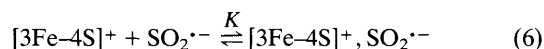
solutions ~ 0.01 M ($1 \text{ M} = 1 \text{ mol dm}^{-3}$) were made up in a Miller-Howe glove box ($\text{O}_2 < 5$ ppm). A 20-fold excess of edta over enzyme was used to complex any free Fe^{2+} . In kinetic runs the variation of [edta] showed no effect on k_{obs} provided $[\text{Fe}^{2+}]$ is corrected for the amount of edta (3–14%) added. Solutions were made up at pH 7.4 using 85 mM HEPES buffer (Sigma Chemicals), and the ionic strength I was adjusted to 0.100 M with NaCl. Solutions of iron(II) ammonium sulphate (BDH, Analar), or iron(II) perchlorate (G. F. Smith, USA) were used as the source of Fe^{2+} . Aconitase concentrations were in the range $(1.5\text{--}2.5) \times 10^5$ M.

Both reactions were studied by stopped-flow spectrophotometry. All solutions were transferred manually from the glove-box to the stopped-flow by syringe.

For the dithionite reduction of inactive $[3\text{Fe-4S}]^+$ aconitase the reaction was monitored at 440 nm. The rate law conforms to the empirical eqn. (4).

$$k_{\text{obs}} = \frac{[\text{S}_2\text{O}_4^{3-}]^{\frac{1}{2}}}{a[\text{S}_2\text{O}_4^{2-}]^{\frac{1}{2}} + b} \quad (4)$$

Thus a plot of $[k_{\text{obs}}]^{-1}$ vs. $[\text{S}_2\text{O}_4^{2-}]^{-\frac{1}{2}}$ is linear, range of $[\text{S}_2\text{O}_4^{2-}]$ employed $(0.16\text{--}8.55) \times 10^{-3}$ M. The behaviour observed is consistent with the mechanism shown in eqns. (5)–(7), where (5) and (6) are rapid. The equilibrium constant K_d has been determined previously and is 1.4×10^{-9} M.⁹ From the present study $K = (2.62 \pm 0.03) \times 10^6 \text{ M}^{-1}$ and $k = 0.44 \pm 0.01 \text{ s}^{-1}$ at 25 °C.



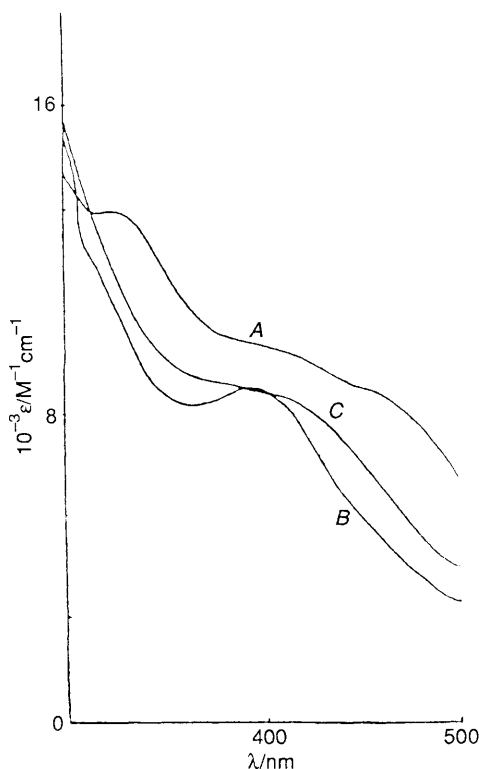


Fig. 1 UV-VIS spectra of aconitase clusters, $[3\text{Fe-4S}]^+$ in 90 mM HEPES (pH 7.5) (A); $[3\text{Fe-4S}]^0$ obtained on reduction of A (B); and $[4\text{Fe-4S}]^{2+}$ after Fe^{2+} incorporation in 90 mM (pH 7.5) (C).⁸ [HEPES = *N*'-(2-hydroxyethyl)piperazine-*N*-ethanesulphonic acid.]

In order to study the reaction with Fe^{2+} , the $\text{S}_2\text{O}_4^{2-}$ reduction was first carried out in the glove-box. Solutions were then transferred to the stopped-flow, and the formation of $[4\text{Fe-4S}]^{2+}$ monitored at 440 nm. The rate law with no sulphate present is as in eqn. (8).

$$\text{Rate} = k_{\text{obs}} [3\text{Fe-4S}^0] \quad (8)$$

where $k_{\text{obs}} = k_0 + k_1[\text{Fe}^{2+}] \quad (9)$

With sulphate present the FeSO_4 complex also participates, see eqn. (10).

$$k_{\text{obs}} = k_0 + k_1[\text{Fe}^{2+}] + k_2[\text{FeSO}_4] \quad (10)$$

Under the condition $[\text{SO}_4^{2-}] \gg [\text{Fe}^{2+}]$ the following dependence see eqn. (11) has been shown to apply, where K is for the formation of FeSO_4 from Fe^{2+} and SO_4^{2-} .

$$\frac{(k_{\text{obs}} - k_0)(1 + K[\text{SO}_4^{2-}])}{[\text{Fe}^{2+}]} = k_1 + k_2K[\text{SO}_4^{2-}] \quad (11)$$

Using a literature value $K = 2.17 \text{ M}^{-1}$,[†] rate constants (25 °C) are $k_1 = 1.99 \pm 0.11 \text{ M}^{-1} \text{ s}^{-1}$ and $k_2 = 269 \pm 12 \text{ M}^{-1} \text{ s}^{-1}$, indicating a much enhanced reaction of neutral FeSO_4 over Fe^{2+} . The presence of charged residues in the cleft is noted. One reason for investigating the effect of SO_4^{2-} relates to the observation that in the crystal structure sulphate is present within the molecule and close to the active site. The function and/or significance of the SO_4^{2-} is not clear. The term $k_0 = (2.83 \pm 0.53) \times 10^{-3} \text{ s}^{-1}$ involves a prior association of Fe^{2+} with enzyme. Since $[4\text{Fe-4S}]^{2+}$ is the product from this step, one possibility is that it arises from inactive $[3\text{Fe-4S}]^0$ which has already become associated with Fe^{2+} .

Investigations are at present being extended to include reactions of other 2+ transition-metal ions with $[3\text{Fe-4S}]^0$, which become incorporated at the Fe_a site. The chemistry of such mixed-metal clusters and nature of the Fe specificity are of interest.

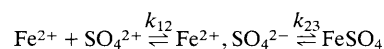
We are grateful to Professor Helmut Beinert for helpful advice with the aconitase isolation procedure, and to the University of Newcastle upon Tyne Research Committee and Henry Lester Trust Ltd for financial support.

Received, 3rd May 1991; Com. 11021241

References

- 1 M. H. Emptage, in *ACS Symposium Series 372: Metal Clusters in Protein*, ed. L. Que, Jr., 1988, p. 343.
- 2 J. F. Morrison, *Biochem. J.*, 1954, **56**, 99.
- 3 B. Fansler and J. M. Lowenstein, *Methods Enzymol.*, 1969, **13**, 26.
- 4 Sr. C. Kennedy, R. Rauner and O. Gawron, *Biochem. Biophys. Res. Commun.*, 1972, **47**, 740.
- 5 M. C. Kennedy, M. H. Emptage, J.-L. Dryer and H. Beinert, *J. Biol. Chem.*, 1983, **258**, 11098.
- 6 A. H. Robbins and C. D. Stout, *Proc. Natl. Acad. Sci., USA*, 1989, **86**, 3639.
- 7 A. H. Robbins and C. D. Stout, *Proteins: Structure, Function and Genetics*, 1989, **5**, 289.
- 8 M. H. Emptage, J.-L. Dryer, M. C. Kennedy and H. Beinert, *J. Biol. Chem.*, 1983, **258**, 11106.
- 9 D. O. Lambeth and G. Palmer, *J. Biol. Chem.*, 1973, **248**, 6095.
- 10 K. Kustin and J. Swinehart, *Prog. Inorg. Chem.*, 1970, **13**, 107.
- 11 M. Eigen and K. Tamm, *Z. Elektrochem.*, 1962, **66**, 107.

† The formation constant K for FeSO_4 was estimated from consideration of the following equilibria,



where $K = K_{12}K_{23} = [\text{FeSO}_4]/[\text{Fe}^{2+}][\text{SO}_4^{2-}]$. Values of $K_{12} = 13 \text{ M}^{-1}$ and $K_{23} = 0.167$ were from refs. 10, 11.